

Useful Byproducts from Cellulosic Wastes of Agriculture and Food Industry—A Critical Appraisal

HIMANISH DAS and SUDHIR KUMAR SINGH

FC Division, Defence Food Research Laboratory, Siddharthanagar, Mysore-570 011, India

Cellulose, an important cell wall polysaccharide, which is replenished constantly in nature by photosynthesis, goes waste in a lion's share in the form of pre-harvest and post-harvest agricultural losses and wastes of food processing industry. These cellulose wastes have an immense potential to be utilized for the production and recovery of several products and ingredients in food application. In this present study, a wide spectrum of researches in the arena of properties of cellulose, hemicellulose and lignin; their degradation; sources and composition of cellulosic and lignocellulosic wastes of agriculture and food industry; present status of converting them into value-added products of food applications; constraints in their conversions and future prospects therein has been reviewed in details. The study has encompassed production of biomass for various utilization and production and recovery of protein and amino acids, carbohydrates, lipids, organic acids, foods & feeds and other miscellaneous products.

Keywords cellulose wastes, value-added products, food application

1. INTRODUCTION

Cellulose is the most abundant material in the universe and makes up more than 50% of the total organic carbon in the biosphere¹ and one third of the vegetation. About 10^{11} tons of cellulose are synthesized annually,² and unlike other natural resources, it is replenishable by photosynthesis.³ The consumption of cellulose is to the tune of 3 folds of steel consumption and almost equal to the quantity of cereal consumption per annum. Its main uses are in the form of wood for construction, furniture, papers, boards, cloths, foods, and feeds. In developing countries, like India, a major portion of the cellulose generated remains unutilized and goes for wastes, namely in the forms of waste from forest resources, wood, paper, and board after human uses, pulp, textile, agriculture, and food industries, etc. Major agriculture, and food industry wastes include straw, leaves, cobs, peels, etc., and damaged agricultural produces due to pre-harvest and post-harvest losses. For every one kg of grains harvested, 1–1.5 kg of straw, cobs, or other residues are generated.⁴ India produces about 127 million tons of fruits and vegetables and 198 million tons of food grains, generating around 200 million

tons of agricultural waste. The post harvest losses of fruits and vegetables in India range from a minimum of 5% to the maximum of 100%.⁵ The area of fruits itself it amounts to 20–30% of the produce, with an annual loss of 3,000 crore of Indian currency.⁶ Overall food wastage for grain products, fruits, and vegetables are 32%, 23%, and 25%, respectively in the USA.⁷ It has been estimated that the dry weight of organic solid wastes annually produced in the USA is 940×10^6 tons, much of which is cellulosic.⁸ The developed countries, like Japan, reuse more than 40% of their urban waste cellulose.⁹ In the present study, properties of cellulose, hemicellulose, lignin and their degradation, the source of cellulosic and lignocellulosic waste of agriculture and food industries, present status and contemporary R and D activities in converting them into value-added products, constraints in their conversions, and future prospects therein have been discussed.

2. PROPERTIES OF CELLULOSE, HEMICELLULOSE, AND LIGNIN

In addition to cellulose, the cellulosic waste materials contain hemicellulose, lignin, extractives, and inorganic compounds.¹⁰ The cellulosic waste from crop residues and other agricultural wastes contain 31–60% cellulose, 11–38% pentosans, and 12–28% lignin. Classically, cellulose is defined as a linear

Address correspondence to Himanish Das, FC Division, Defence Food Research Laboratory, Siddharthanagar, Mysore-570 011, India. E-mail: dfoodlab@sanchamat.in

polymer of anhydroglucose units linked at C₁ and C₄ atoms by a β -glucoside bond. The degree of polymerisation in cellulose molecule is found to be a considerably varying one.¹¹⁻¹³ Various models of cellulose structure have been proposed by Chang¹⁴ and other authors. Lignin is a complex of 3-D polymer of phenolic and enolic origin, whereas hemicelluloses comprise of cellulose and polyuronide hemicellulose.^{15,16} Overall, lignins are rather heterogeneous and are branched polymers of indefinite structure and size.¹⁷ Cellulose is insoluble in water, but soluble in a number of solvents, including concentrated acids and inorganic solvent solutions. Constraints in cellulose degradation have been reviewed by several authors.^{18,19} Cellulose itself is among the least degradable natural polymers due to its high molecular weight, high degree of structural order, insolubility, and low surface area. Its association with lignin and hemicellulose makes the plant materials more stable to chemicals and enzymes. Hence, extensive pretreatment is often required to encourage degradability of the natural cellulose.²⁰⁻²⁴ Four types of degradation have been characterized, namely hydrolytic, oxidative, microbial, and mechanical degradation.²⁵ The addition of NaOH prior to thermal pretreatment often enhances the degradability of wastes by disrupting lignin and lignocellulose structures, which might physically restrict the extracellular hydrolyzing enzymes of microorganisms.^{26,27} Pretreatment of celluloses is done to increase the rate and the extent of microbial digestion.²⁸⁻³⁴ Also, enzyme action on cellulose releases chemically bound lignin. A synergistic effect of various cellulolytic microorganisms³⁵⁻³⁹ and that of the combination of the purified enzymes from culture filtrate of *Trichoderma viride* on the hydrolysis of cellulose⁴⁰ are reported. Alkali or petrochemical treatment, electron irradiation, and ball milling to a fine particle size (70-100 μ), etc., increase the enzyme activity and biodegradability of cellulose by various fungi. Cellulose can be pretreated and swollen by some strongly electrolytic solvents, acids, etc.⁴¹ According to Desai and Betarbet,⁴² cellulose degradation is caused by cellulase secreting fungi, bacteria, and actinomycetes. The cellulase is a complex enzyme, which consists of β -1,4-endoglucanase and β -1,4-exoglucanase. A third enzyme β -glucosidase (Cellobiase) is always associated with the two glucanases and is essential for the complete hydrolysis of cellulose to glucose.⁴³ The cellulolytic bacteria most studied are *Cellvibrio (Pseudomonas) fulvus* and *Pseudomonas fluorescens* strain.^{44,45} Few other cellulolytic organisms studied are *Coniophora cerebella* and *Sporotrichium pulverulentum* under Basidiomycetes; *Trichoderma viride*,⁴⁶ *T. koningii*,^{46,47-52} *Penicillium funiculosum*^{53,54} and *Fusarium solani*^{57,55,56} under Ascomycetes and Deuteromycetes; thermophilic fungi: *Chaetomium thermophile var. dissitum*, etc.

3. SOURCES

From the agricultural farms, plantations, and orchards, the cellulosic residues generated in the form of stalks, straws, stems, leaves, cobs, chaffs, bunches, stumps, and stubbles, damaged grains, fruits, vegetables, etc. The major cellulosic wastes from

Table 1 Cellulose content and composition (g/100 g of dry matter)

Cellulosic wastes	Cellulose	Lignin	Hemicelluloses
1. Agricultural residues			
Barley straw ⁵⁷	44	7	27
Oat Straw ⁵⁷	41	11	16
Oat Straw ⁵⁸	42.8	—	—
Rice Straw ⁵⁷	33	7	26
Rice Straw ¹⁰	32.1	—	—
Wheat Straw ⁵⁷	39	10	36
Wheat Straw ¹⁰	30.5	—	—
Sorghum Straw ¹⁷	31	11	30
Cottonseed Hulls ⁵⁷	59	13	15
Sugarcane Bagasse ⁵⁷	40	13	29
Entire Bagasse ¹⁰	46	—	—
Fibre Bagasse ¹⁰	56.6	—	—
Pith Bagasse ¹⁰	55.4	—	—
2. Fruits & Vegetables ⁵⁹			
Apples	2.9	Trace	5.8
Banana	1.3	0.93	3.83
Lemons	—	35	—
Oranges	—	14	—
Pineapples	—	7.64	—
Strawberries	3.6	8.4	10
Potato	1.2	Trace	9.2
Carrot	12.9	Trace	19
Cauliflower	13.4	Trace	13
Cabbage	8.9	4.3	26
Tomato	9.1	5.3	11
Peas	14	2	36
3. Seeds ⁶⁰			
Barley	5.3	—	—
Corn	2.4	—	—
Wheat	2.1	—	—
Oat	11.9	—	—
Grain Sorghum	2.7	—	2.5
Peanuts	2.8	—	2.5

food industries include damaged fruits, vegetables and grains, and post-processing residues, like skins, peels, seeds, leaves, bunches, husks, bagasse, vinasse, pomace, etc. These vary in cellulose content and composition as described in Table 1.

4. CONVERSIONS OF CELLULOSICS AND R AND D ACTIVITIES

Cellulose is a great reservoir of energy in the world. Cellulosics generated from agriculture and food processing industries have been given great attention by various workers because they add value to products. Some of these are biomass production, production and recovery of protein, amino acids, etc., carbohydrates production, acid production, lipid production, production of foods and feeds, and others.

4.A. Biomass Production

Microorganisms are grown on cellulosic substrates for their biomass production in view of their use as or in foods, feeds, the production of enzymes, single cell protein (SCP), amino acids, lipid, carbohydrates, and organic acids. A large number of fungi have been screened for their ability to grow on cellulose.⁶¹

Lignocellulosic and other food and agricultural wastes have been utilized for the production of mushrooms.⁶² Sugarcane leaf and bagasse are obtained in huge mass from sugarcane field and sugar industry. Sugarcane bagasse and bagasse pith both pre-treated with NaOH solution have been used separately for the growth of *Cellulomonas* spp.^{63,64} A biomass production by growing a defined mixed culture of *Trichoderma viride* and *C. utilis* on pretreated bagasse has been attempted.⁶⁵ Sugarcane (*Saccharum sinensis*) bagasse or tobacco (*Nicotiana tabacum*) stalks, when fortified with $(\text{NH}_4)_2\text{SO}_4$ and KH_2PO_4 , produces a fresh mushroom, *Pleurotus sajor-caju*.⁶⁶ Some were able to produce bacterial biomass of *Cellulomonas* sp. and *Thermomonospora fusca* on physically and chemically pretreated cellulosic wastes, like bagasse and straw.^{67,68} Cellulosic wastes from citrus processing⁶⁹ and tea production unit⁷⁰ have been found to support the biomass production of edible fungi basidiomycetes. A good production of edible mushroom, such as *Agaricus bisporus* and *Morchella crassipes*, has been noticed on lime waste.⁷¹ In another study, successful utilization of date wastes, such as substrates has been indicated for the production of yeast (*Saccharomyces cerevisiae*) biomass.⁷² Sturion and Oetterer⁷³ have conducted studies on waste banana leaf and prescribed it as a more efficient substrate than wheat straw for the cultivation of edible fungi, such as *Pleurotus* sp. *subsp. florida*, *Pl. obstromoseus* and *Pl. sajor-caju*. Soybean wastes from the processing of soy-milk and tofu have been utilized for the growth of various edible fungi, namely *Coriolus versicolor*, *Tricholoma lobayense* T-2, *Lycophyllum shimeji* M-46 *Morchella elata* M-26, *Pl. sajor-caju*, PI-27, and *Volvariella volvacea* V.34.⁷⁴ These are all potential producers of high value metabolites, including nutraceuticals. Cellulolytic *Aspergillus* sp. strains VK PMF 559 isolated from a water treatment plant can convert 80% of cellulose in corn cobs into its biomass.⁷⁵ *Penicillium expansum* is able to yield a biomass in a mineral medium containing acid-treated rice husk.⁷⁶ An early rapid proliferation of yeast in food waste containing rice and vegetable residues has been observed,⁷⁷ but has been described as an insignificant activity by Nakasaki and colleagues.^{78,79} The cellulose rich hulls, when treated with NaOH and heat, have produced a black liquor, which has been used as a substrate for the growth of 5 species of *Aspergillus*, 3 spp. of *Paecilomyces*, 2 spp. of *Penicillium*, 1 spp. each of *Alternaria*, *Trichoderma*, *Chaetomium*, and *Actinopolyspora*.⁸⁰

4.B. Recovery and Production of Protein and Amino Acids

Single cell protein (SCP) production from cellulose has been preliminarily evaluated, suggested, or reviewed by several authors.^{67,81,82} Some agricultural by-products, such as cereal straws are in use for microbial protein production, but still more can be used (especially if alkali treated) without any harm to the live weight gain of the animal fed with this.^{83,84} A method for the production of nutritive protein by growing a cellulase producing microorganism and *Alcaligenes faecalis* on delignified cellulosic materials, like bagasse, rice straw, Johnson grass, Prairie grass,

oat straw, wheat straw, sorghum bagasse, corncobs, and cottonseed hulls has been described.⁸⁵ Barley straw has been converted by *Trichoderma viride* alone or a mixed culture of *Trichoderma viride* and *C. utilis* into a product containing 21–26% protein. Other cellulosic wastes can also be utilized.⁸⁶ The interest of the bioconversion of lignocellulosic material for protein enrichment of animal feeds using SSF has been highlighted.⁸⁷ A dilute acid hydrolysis without separation of the products (SCP) as they are formed is commonly used when straw is the substrate. An alternative way to produce SCP from cellulose materials after their enzymatic and microbial degradation to glucose has been proposed by Brown and Fitzpatrick⁸⁸ and supported by Kamikubo and colleagues.⁸⁹ Enzymatic hydrolysate of cellulosic wastes of tea concentrate has also been used to produce SCP.⁹⁰ Sorghum husk, millet husk, and banana stem digested with 0.6N H_2SO_4 , have been used in a culture medium to grow *Penicillium expansum*, in order to produce a considerable quantity of histidine and lysine amino acids.⁹¹ This organism has also produced SCP (37–62%) on a medium containing similarly treated rice husk.⁷⁶ The cells of a photosynthetic bacterium, *Rhodospseudomonas gelatinosa*, grown on wheat bran infusion, contain significant quantities of protein and vitamins. Moo-Young⁹² has proposed several fermentation processes for the production of SCP. A novel multistage fermentation process has been proposed for the production of SCP (*Chaetomium cellulolyticum*) to be used as an animal feed supplement. The mycelial mass is found to be nutritious, digestible, and non-toxic. This SCP resembles highly with fodder yeast and soy meal in terms of essential amino acid composition, but, as per FAO reference, this is deficient in sulfur containing amino acids.⁹³ The stillage residues from the ground high lysine corn used for alcohol fermentation can be fractionated into distiller's grain, centrifuged solids, and stillage soluble. The centrifuged solids have the highest protein content (37–49%), followed by distiller's grain (31–35%). The latter has potential for use in human food and non-ruminant feed. On the other hand, the stillage soluble is found to have a high lysine content.⁹⁴ A process has been developed for SCP production, upto 60% crude protein by *Cellulomonas* spp. grown on sugarcane bagasse.⁹⁵ The treatment of bagasse with 1% NaOH yields a maximum protein production with a mixed culture of *Cellulomonas* spp.-*Bacillus subtilis*.⁹⁶ A remarkable work has been carried out on pretreated Egyptian sugarcane bagasse.⁶⁵ The bagasse pretreated with alkali plus γ -irradiation, acid plus γ -irradiation, and NaClO_3 enables a mixed culture of *Trichoderma viride* and *Candida utilis* to produce a biomass containing 35.5% protein and all the essential and non-essential amino acids in accordance with FAO reference. A growth of *Aspergillus* spp. strain VK PMF 559 on corn cobs⁷⁵ and Basidiomycetes on citrus wastes⁶⁹ or higher Basidiomycetes (*Cerrena unicolor*, *Coriolus hirsutus*, *Pleurotus ostreatus* IBK-191) on tea waste⁷⁰ has produced a biomass containing a varied quantity of protein. Lee and Kim⁹⁷ have extracted proteins from soymilk residue using enzymes from *Asp. oryzae*. Various fruit and vegetable waste from the aubergene, tomato, grape, apple, cabbage, carrot, beet root, and watermelon are the substrates for lysine production by

*Brevibacterium spp.*⁹⁸ A protein enrichment has been obtained by growing *Aspergillus niger* on mango peel, orange peel, green and over-ripe banana, and carrot waste in SSF.⁹⁹⁻¹⁰¹ The protein content of apple pomace to be used as an animal feed has been enriched by SSF and submerged fermentation by various co-culture of cellulolytic fungi and yeasts, out of which a co-culture of *Candida utilis* and *Asp. niger*, was the best performer in increasing the protein content of dried pomace and pectin extracted apple pomace to 20% and 17% respectively under SSF.¹⁰² Similarly native and mutant strains of *Trichoderma aureoviride* have been used in a solid state column fermentation of leached sugar beet pulp waste for protein enrichment upto 40% of total dry weight.¹⁰³ Edible fungi basidiomycetes grown on citrus waste can produce biomass with high protein content of about 18-19%.⁶⁹ *Aspergillus niger* TMF-15 is able to convert 90-92% of cellulose in a mixed substrate of extracted grape waste and pressed apple pulp into a product having a higher protein content (35%) than those obtained from *Penicillium funiculosum* 515 and *Myrothecium verrucaria* 9095.¹⁰⁴ An enrichment of protein upto 32-34% has been observed by growing *Chaetomium globosum* and *Sporotrichium pulverulentum* on kinnow—mandarin wastes of peel, pulp, and seeds.¹⁰⁵ SCP is produced from banana wastes, employing *Pichia spartinae* and date wastes by a mixed culture of *Saccharomyces cerevisiae*, *Candida utilis*, and *Saccharomyces rouxii*. A five-fold improvement in the protein content of cassava meal has been reported.¹⁰⁶ Sun dried tapioca fibrous residue¹⁰⁷ and soybean waste residue¹⁰⁸ can also be used for SCP generation. Various fungi have been investigated for their growth on coffee processing waste in ICAITI, Guatemala.^{109,110} *Trichoderma harzianum* produces dry solids containing about 56% protein from coffee processing wastes. A trial experiment has been conducted by growing ten strains of yeasts on Egyptian vinasse with the possibility of successful recycling of the waste for microbial protein production.¹¹¹ Various substrates are used by various researchers for the production of biomass having enriched protein content (Table 2). Cabbage can be utilized economically by enriching its protein content by native commensal microflora.¹¹² Cocultures of cellulolytic and non-cellulolytic microbes have demonstrated higher yield and productivity than the monocultures.¹²² It is because cellulases from cellulolytic species catalyze cellulose to glucose, which is, in turn, fermented by non-cellulolytic microorganism. A wide range of cellulases obtained from *Trichoderma spp.* has been well-characterized.¹²³ The SCP produced by *C. utilis* grown on acid and enzymatic hydrolysates of waste rice straw should be used as protein supplement and not as a major diet component from a nutritional safety point of view.¹²⁴ The fungal mass grown on various crop residues is found as a safe and acceptable food product, as implied by toxicity trials and food textural studies.¹²¹

4.C. Production of Carbohydrates

Pretreatments of the cellulolytic waste from crop residues and other agriculture wastes with acids,¹²⁵ alkali,¹²⁶ or enzymes¹²⁷

are effective to release a variety of fermentable sugars. The sun dried tapioca fibrous waste residue can be utilized for the production of glucose syrup, high fructose syrup, and confectioner's syrup.¹⁰⁷ It has been strongly supported by a study on the conversion of cellulose slurries to glucose syrup.¹²⁸ Cellulosic cassava waste saccharified with a culture filtrate from *Trichoderma viride* and a soil Basidiomycetes, together with commercial amylases, enables an enhanced sugar formation. A syrup containing a high concentration of xylose and other sugars can be produced by a two phase acid hydrolysis of hemicellulose-rich plant materials, viz., are leaves and stems of sorghum, maize, or wheat.¹²⁹ A xylose syrup has been generated from acid-hydrolysed xylan-rich cottonseed hulls.¹³⁰ A large-scale enzymatic depolymerization of xylan extracted from corncob meals has yielded various sugars, like xylobiose, xylotriose, and arabinose oxo-oligosaccharides.¹³¹ The enzyme used is endo-1,4- β -xylanase, obtained from *Clostridium thermolacticum* DSM 291. A medium containing straw, microcrystalline cellulose and xylose, has been utilized to produce endoglucanases and xylanases (by *Alletheria terrestris*), which have been employed to hydrolyze cellulose and hemicellulose present in fruit canneries wastes to yield monosaccharides predominantly glucose.¹³² Toyama and Ogawa¹³³ have been able to produce 10% sugar syrup from a substrate made up of rice straw and bagasse delignified by boiling with 1% NaOH and saccharified by a commercial cellulase preparation (1% Meicelase) from *Trichoderma viride* at 25% substrate concentration. Conversion of ignocellulosic residue of corn-cobs or alkali- and heat-treated hulls into pentoses have been studied separately.^{80,134} Fungal enzymes cellulases and hemicellulases have been used to saccharify agrowaste.¹³⁵ Enzymes from *Aureobasidium spp.* strain NRRL Y-2311-1 to saccharify corn fibres¹³⁶ and other enzymes¹³⁷ have been used to saccharify coffee bean solid waste. In India, efforts have been directed to utilize mango-processing waste for the recovery of starch.¹³⁸ The microbial production of polysaccharides from agricultural wastes or a mixture of pure chemical and agricultural wastes has been studied.¹³⁹ Grape skin pulp extract¹⁴⁰ and carob pod¹⁴¹ are used as substrates for the production of pullulan by *Aureobasidium pullulan* NRRL-Y 6220. Xanthan gum has been produced by *Xanthomonas campestris* ATCC 13951 from four types of citrus waste fractions viz., whole citrus waste from oranges, lemons and grape fruits, pectic extracts, hemicellulosic extracts, and cellulosic extracts. The biodegradability of hemicellulosic and cellulosic wastes is found to be lower than that of the other two fractions.^{142,143} An alkaline extraction of xyloglucan from depectinized apple pomace¹⁴⁴ and production of cyclodextrin from rice bran¹⁴⁵ have also been reported. Applications of microbial polysaccharides¹⁴⁶ and xanthan gum in foods¹⁴⁷ have been summarized.

4.D. Production of Lipid

Fat production by yeast and other microorganisms has been adequately highlighted.^{148,149} A correlation has been established between the possession of the enzyme, ATP-citrate lyase, and

Table 2 Protein content of biomass

Microbial biomass	Substrates	Composition (g/100 g dry wt.)		
		N ₂	Protein	Fat
A. Bacterial biomass				
<i>Cellulomonas</i> sp. ⁶³	Bagasse	9.3	58	—
<i>Cellulomonas</i> sp. & <i>Alcaligene</i> sp. ²⁹	Bagasse	14	87	8
<i>Cellulomonas</i> sp. & <i>Alcaligene faecalis</i>	Alkali treated rye grass	—	6.8	—
	NH ₃ treated rye grass	—	9.5	—
<i>Cellulomonas</i> sp. & <i>Candida utilis</i> ¹¹³	Barley straw	9.3	10.6	58-66
<i>Bacillus</i> sp. ¹¹⁴	Rice hulls	4.5	28	—
<i>Pseudomonas</i> sp. & <i>Chaetomium cellulolyticum</i> ¹¹⁵	4% NaOH treated rice straw	—	19	—
	4% NaOH treated corn stover	—	20-24	—
	3% NH ₃ treated corn stover	—	17	—
B. Filamentous fungi				
<i>Aspergillus fumigatus</i> I 21 ^{116,117}	Cassava extract	40	31.5	12.2
	Cassava	37	27	—
<i>Chaetomium cellulolyticum</i> ¹¹⁸	Crop residues	45	—	10
<i>Fusarium moniliforme</i> ¹¹⁹	Carob extract	43	30	5
<i>Rhizopus chinensis</i> ¹²⁰	Cassava extract	49	37	—
<i>Sporotrichum album</i> ¹¹⁶	Cassava extract	64	54	6-12
<i>Aurobasidium pullulans</i> ¹²¹	Acid-treated and NH ₃ -neutralized straw	—	14	—
<i>Candida utilis</i> ¹²¹		—	12.4	—
<i>Trichoderma viride</i> ¹²¹		—	10.9	—
<i>Fusarium semitectum</i> ¹²²	Oat husk	—	11	—
	Sugarcane bagasse	—	5.6	—
	Cocoa shell	—	5.9	—
	Spent hops	—	5.9	—

the ability of yeast to accumulate more than 20% of its biomass as a lipid.¹⁵⁰ Fat production by red yeast, *Rhodotorula gracilis*, seems very promising. The only bacterium, which has been reported as a producer of significant amount of triacylglycerol, is *Arthrobacter* AK 19.^{151,152} Date extract and rice hull hydrolysates have been considered as a substrate for the production of microbial oils by *Penicillium soppi* and *Lipomyces* spp., respectively. A few specialized cellulolytic fungi appear to be non-oleaginous, but cellulose, after its hydrolysis, can be a suitable form of fermentable carbohydrate for them. The estimated cost per ton of generated fermentable carbohydrate is not too high to use cellulose as a substrate for microbial oil production. Some yeasts grow on hydrolysates and oxidates of cellulosic peat for production of both protein and oil.^{153,154} One strain each of *R. gracilis* and *Trichoderma cutaneum* is found to be oleaginous with the lipid content of the culture being 21.9% and 26.1% on yeast dry matter, respectively. When grown on a synthetic medium and a carrot juice containing medium, *R. gracilis* has produced cells containing 25% and 17-20% lipid and *T. cutaneum* with 35.5% and 21-25% lipid.^{155,156} An extraction of oil from olive pomace, apple pips, and mango processing wastes has been reported.^{157,158}

4.E. Production of Organic Acids

Cellulosic wastes of agriculture and food processing can also be made use of for the production of various acids having a potential use in the food and chemical industries. Vine-

gar can be prepared from citrus peel or molasses,¹⁵⁹ mango processing waste,¹³⁸ fermentation of cellulosic waste of potato starch industry by *Fusarium oxysporum* 841,¹⁶⁰ and waste from pineapple juice.^{161,162} An interesting conversion of cellulose to acetic acid by a new sewage isolate of *Aceto-vibrio* spp has been proposed.¹⁶³ *Asp. niger* has been employed by a few researchers for citric acid production from a variety of cellulosic substrates.^{164,165} The pineapple peel, apple pomace, wheat bran, and rice bran are utilized as substrates for the production of citric acid in SSF by *Aspergillus foetidus* ACM 3996. Pineapple peel is found to be a better substrate than others, producing the highest yield at 16.1 g citric acid/100 g dry peel.¹⁶⁶ Hang and Woodams¹⁶⁷⁻¹⁷⁰ have been able to produce citric acid successfully from apple pomace, grape pomace, kiwi fruit, and pineapple peel in a SSF. Waste from the preparation of guava jelly has enough cellulosic material to be used for citric acid production by microorganisms.¹⁷¹ Citric acid has been generated from dewatering of orange peel by using 3 strains of *Aspergillus niger*. The greatest yield of 30 g citric acid per kg press liquor is produced by strain NRRL-599.¹⁷² By cultivating *Asp. niger* CFTRI 30 on dry coffee husk, potentiality of production of citric acid has been commercially demonstrated.¹⁷³ *Asp. niger* NRRL 2270 and *Rhizopus oryzae* NRRL-395 are found to generate citric acid and lactic acid with 36% and 58% yield, respectively.¹⁷⁴ Cocoa juice, a waste from cocoa bean fermentation, has supported the growth of *Candida lipolytica* ATCC 8661 and *Asp. niger* ATCC 10581 to produce citric acid.¹⁷⁵ Sugarcane press mud is a novel and inexpensive substrate for

citric acid production in SSF by *Lactobacillus casei* subsp. *casei* CFTRI 2022, *Lb. velveticus* CFTRI 2026, and *Streptococcus thermophilus* CFTRI 2034.¹⁷⁶ Cellulosic materials, such as comcobs, corn stalks, cottonseed hulls, and straw can also be used for the production of lactic acid.¹⁷⁷ In a lactic acid fermentation of the cellulosic materials, the recovery of lactic acid from the crude broths and presence of lignin pose additional problems for their further conversion.¹⁷⁸ The manufacture of tartaric acid from grape pomace¹⁷⁹ and oxalic acid from apple pomace¹⁵⁸ has been reported. Cassava bagasse, a starch-rich lignocellulosic residue, has been established as a sole carbon source to produce fumaric acid by several *Rhizopus* strains in submerged fermentation. The strain, *Rhizopus formosa* MUCL 28422, has been reported as the best producer.¹⁸⁰ Fumaric acid, due to its non-toxic and non-hygroscopic properties, is used as an acidulant in food and beverages. Propionibacterium or mixed bacterial culture is able to ferment lignocelluloses to produce propionic and or acetic acid.^{181,182} Dynatech workers have successfully fermented aquatic weed to produce organic acids in a high yield with 43–45% conversion of algal species and with the production of acetic acid up to 40–60% of total acids.¹⁸³

4.F. Production of Foods and Feeds

Ruminants retain cellulose for a long time in their alimentary canal, the micro-flora within which aid in digestion. The cereal straw has long been used as animal feed. The shortcomings of straw as a feed are its low digestibility, low protein content, poor palatability, and bulkiness. The digestibility of lignocellulosic straw is generally inversely related to the amount of lignin present.^{184–187} The digestibility of waste plant materials can be improved by acid/alkali treatment,^{188–190} steaming at super atmospheric pressure,¹⁹¹ irradiation,¹⁹² or microbiological treatment.^{30,193–197} The physico-chemical mechanism of pretreatment for increased digestibility of lignocelluloses have been elucidated.^{58,193,198} A number of oxidizing agents have been used on sugarcane bagasse for this purpose.¹⁹⁹ The dioxane treatment has improved the digestibility of rye grass straw and rice straw, but not that of sugarcane bagasse.²⁰⁰ The method of converting sugarcane stalks and tops by Bishop,²⁰¹ sugarcane pith by Miller and Lauric,²⁰² and leaves, tops, and short top pieces by Worden²⁰³ have been patented. An extrusion process of tomato pomace has been described, and the product has been evaluated for its composition.²⁰⁴ *Kloceckera apiculata* and *Candida utilis* can transform apple pomace into an improved stock-feed by SSF.²⁰⁵ Use of fruit pomace for the manufacture of functional drinks and sour fruit beverages has been prescribed.^{206,207} An animal feed²⁰⁸ and a sauce²⁰⁹ from apple pomace have been produced and evaluated. The effective utilization of carrot pomace in bread, cake, dressings, and pickles has been studied in details.^{210,211} A protein-rich fibrous foodstuff called germinated barley has been made from brewers' spent grain. This product contains glutamine-rich protein and the dietary fibres

of cellulose, hemicellulose, and lignin.²¹² Wu⁹⁴ has shown that distillers' grain, a fraction of stillage residue of alcohol fermentation of ground high lysine corn, has a potential use in human food and non-ruminant animal feed because of its high protein quality. Likewise, a food product for human beings has been developed from distillers' spent cereal grains and solubles.²¹³ A silage preparation technique has been evolved from barley shochu distillery by-product and juice residues of mandarin oranges and carrots.²¹⁴ Coffee pulp, a huge by-product of the coffee processing unit, can be safely used as feed, if the caffeine, tannins, and polyphenol are removed. Several caffeine degrading microorganisms belonging to genera *Aspergillus*, *Penicillium*, *Trichoderma*, *Fusarium*, and *Humicola* have been isolated. It has been found that 5 strains of *Aspergillus* spp. and 2 strains of *Penicillium* spp. can degrade 100% of the caffeine in the liquid medium.²¹⁵ Mahadevaswamy and Venkataraman²¹⁶ have enlightened an integrated approach of utilization of mango processing waste. The effluent of biogas production from mango processing waste has been utilized for the production of fresh water fishes, like major carp, rohu (*Labeo rohita*), and common carp (*Cyprinus carpio*). Cull grape fruit and Jerusalem artichokes have been examined as potential feed-stocks,¹⁷⁷ and the nutritional and toxicological evaluation of wild apricot pomace has been carried out.²¹⁷

4.G. Production of Miscellaneous Products

Several other food ingredients or additives, namely pectin, edible fibre, pigment, flavouring, antioxidant, etc., which have potential application, can be generated from various cellulosic wastes. The utilization of cassava fibrous residue²¹⁸ and wastes from the processing of apple,^{219,220} for value-added products, has been widely reported. The aspect of pectin production from the fruit processing wastes has been reviewed by Sudhakar and Maini.²²¹ Apple pomace and wastes from the processing of peas, peaches, apricots, and other fruits are found to be the good sources of pectin production.²²² The extraction and production of pectin from apple pomace,^{223,224} apple wastes,²²⁵ citrus wastes,²²⁶ orange peel,²²⁷ and mango wastes^{228,229} have been carried out successfully. Food fibre is another ingredient studied by few researchers. The dietary fibre content of pear, kiwi,²³⁰ and grape pomace²³¹ has been described. In India, the utilization of mango processing wastes for the recovery of valuable chemicals, such as pectin, pigments, tannins, and cocoa butter substitutes has been directed. A comprehensive review of Kennedy's research¹⁵⁸ has suggested the potential development of food additives, flavourings, anti-browning agents, etc. from kiwi fruit wastes and the manufacture of pectin, aroma compound, fibre, etc. from apple pomace. The antioxidant activity of potato peel waste or red grape pomace and peel has been reported.^{232–234} Also, a red pigment has been obtained from red grape pomace.²³⁵ An algae, *Chlamydomonas reinhardtii*, is able to produce biomass and photosynthetic pigments when grown on vinasse, a waste from beet and cane fermentation.²³⁶ The

apple industry wastes can also be utilized for the production of biogenous thermoplastic materials.²³⁷ The lignin present in cellulosic waste can also be oxidized by chemical, microbial, or enzymatic processes to produce a flavour compound, vanillin.²³⁸ The banana processing waste of about 8.30 lakh tons generated annually in India can be utilized for the generation of furfural and pectin. The presence of waxes, including triterpenoids in dried apple pomace and its depectinised residue, has been indicated.²³⁹

5. CONSTRAINTS IN CELLULOSE CONVERSIONS AND PROSPECTS

Bottlenecks on the economic utilization of cellulosic wastes of agriculture and food processing are chiefly reported as a lack of rapid enzymatic hydrolysis of cellulose, lack of rapid microbial growth on celluloses, lack of cellulases with high catalytic power, high cost of pre-treatment, and presence or association of other substances with them. The problem with all cellulolytic bacteria is that the in-vitro (cell free) system has a high degree of activity on soluble cellulose derivatives, but very low activity, if any, towards native cellulose. Microbial growth and cellulase production are influenced by various factors, like medium, pH of medium, inoculum, environmental factors, etc.²⁴⁰ The lignin bound to celluloses inhibits microbial attack and enzyme action on celluloses. Martinez and his colleagues²⁴¹ have been able to fractionate residual lignocellulosics from almond shells by dilute acid pre-hydrolysis and alkaline extraction. There are other approaches to the utilization of cellulosic wastes without a separate step of hydrolysis by acid or enzyme. If technically feasible, the direct use of polysaccharide is preferred.²⁴²⁻²⁴⁴ Although the microbial degradation of lignin has been reviewed by several authors,²⁴⁵⁻²⁴⁷ lignin is found to inhibit the breakdown of cellulose by enzymes, acids, or microorganisms.²⁴⁸ Processes in which acids are used to catalyze cellulose hydrolysis are not commercially attractive.²⁴⁹⁻²⁵² This is because the cost of sugar production is high and the yields are typically low. Basically, cellulose is more chemically stable than starch, therefore, harsher acid hydrolysis conditions or more potent enzyme solutions are required for its hydrolysis.²⁴ In Japan, cellulases from *A. niger* and *T. viride* are produced commercially at a rate of 45 tons per year representing an economic output of 170×10^6 yen per year. Presently both the exorbitantly high cost and the limited efficiency of known cellulase preparation make many potential applications non-profitable. The cost of cellulase enzymes encompasses about 60% of total cost of saccharification,²⁵³ therefore, the reuse of the enzyme, as well as its stability, is essentially important. To improve process efficiency, one can search for more active cellulases or develop cheap methods for pretreatment of the substrates. The attention of some workers has been paid onto hemicellulases, pentosanases, and xylanases. The manufacturing cost of SCP is very much dependent upon the cost of carbon and energy source to the producer of SCP products. The raw material cost of bagasse ranges from 17 to 26% of the total cost

of SCP production. On the basis of the work of Abbott²⁵⁴ and Wang,²⁵⁵ protein generated by bacteria grown on cellulose is much cheaper than many other conventional proteins from beef, chicken, eggs, fish, cheese, etc. The cost associated with the collection and transportation of wastes will be excessive, if SCP production plants are far away from the waste generation site. An economic feasibility study on production of animal feed protein from lignocellulosic wastes pretreated with chemicals, enzymes, microorganisms, or a direct bioconversion into protein-enriched fermented fodder and sugar syrup has been extensively carried out in a research programme in KISR.²⁵⁶ Ratledge²⁵⁷ has argued that SCO could be a more valuable product than SCP, as the oil would be several times higher in value than animal protein. Any future economic utilization of celluloses should be based on their current exploitation pattern. Economies derive from the carbohydrate substrate for fermentation from the hydrolysis of crops and forest residues, rather than from grains or other high grade food materials.^{258,259} That is why celluloses have received a great attention as possible feedstocks on the assumption that raw material costs from agriculture and lumber waste are negligible⁴.

6. CONCLUSION

Cellulosics can be utilized for production or recovery of some value-added ingredients or products of various usages, providing monetary benefit to the firms. The Government Regulations on pollution and environment protection, pressure from social groups, and threat from politicians also lay emphasis on the utilization of cellulosic wastes of agriculture and food industry origin. The utilization of cellulose, a replenishable resource, can be transformed into a lucrative and profitable business, if future research in this area is aimed at overcoming or relieving successfully overcoming the constraints underneath it.

REFERENCES

- [1] Lehninger, A.L. 1970. The molecular basis of cell structure and function. In *Biochemistry*. Worth Publishers Inc., New York.
- [2] Ratledge, C. 1977. Fermentation substrates. *Annu. Rep. Ferment. Processes*, 1:49-71.
- [3] Soltes, E.J. 1980. Thermal conversion of lignocellulosics: Retrospect and prospect. Presented at the *Inter-American Seminar/Workshop on materials for the future: Renewable organic resources for industrial materials*. Kingston, Jamaica. 18-21.
- [4] Blanch, H.W. and Sciamanna, A. 1980. Composition of cellulosic raw materials for sugar production. *Alcohol Fuel Process RID Newsletter*. Winter Quarter, USDOE Solar Energy Research Institute, Golden Co., 69.
- [5] Ashok, B.S. 1998. New policy to woo corporate investments. *The Financial Express*. India. 11.
- [6] Food Digest. AFST (1). Mysore, India, vol. 18: no.4, Oct-Dec 1995.
- [7] Food Digest. AFST (1). Mysore, India, vol. 20: no.3, July-Sept 1997.
- [8] Humphrey, A.E. 1975. Production outlook and technical feasibility of SCP. In *Single Cell Protein II*. Tannenbaum, S.S., and Wang, D.I.C., Eds. pp. 1-23. MIT Press, Cambridge, USA.

- [9] Personal Communication. Dunlap, C.E. Louisiana State University. T.C. Purcell. Bureau of solid waste management. March 25, 1969.
- [10] Virkola, N.E. 1975. In *Symposium on enzymatic hydrolysis of cellulose*, p. 23. Bailey, M., Enari, T.-M., and Linko, M., Eds. SITRA, Aulanko, Finland.
- [11] Cowling, E.B. and Brown, W. 1969. Cellulases and their applications. In *Advances in chemistry series*, vol. 95, p. 152. Gould, R.F., Ed. American Chemical Soc. Publications, Washington, D.C.
- [12] Emert, G.H., Gum, E.K., Lang, J.A., Liu, T.H., and Brown, R.D., Jr. 1974. *Food related enzymes*. In *Advances in chem. ser.*, vol. 126, p. 79. Gould, R.F., Ed. Amer. Chem. Soc. Publications, Washington, D.C.
- [13] Sihtola, H. and Neimo, L. 1975. In *Symposium on enzymatic hydrolysis of cellulose*, p. 9. Bailey, M., Enari, T.-M., and Linko, M., Eds. SITRA, Aulanko, Finland.
- [14] Chang, M. 1971. Folding chain model and annealing of cellulose. *J. Polymer Sci. C-36*:353.
- [15] Norman, A.G. 1954. Noncellulosic carbohydrates. In *Cellulose and cellulose derivatives*, Part I, p. 459. Emil, O., Spurlin, M.H., and Graffin, W., Eds. Interscience Publishers Inc., New York.
- [16] Whistler, R.L. and Richards, E.L. 1970. In *Hemicelluloses, the carbohydrates: Chemistry and biochemistry*, p. 447. Pigman, W. and Harton, D., Eds. Academic Press, New York.
- [17] Goring, D.A.I. 1971. Polymer properties of lignin and lignin derivatives. In *Lignins, occurrence, formation, structure and reactions*, pp. 695-768. Sarkanen, K.V. and Ludwig, C.H., Eds. Wiley-Interscience, New York.
- [18] Cowling, E.B. 1975. Physical and chemical constraints in the hydrolysis of cellulose and lignocellulose materials. *Biotechnol. Bioeng. Symp.*, 5:163-181.
- [19] Fan, L.T., Lee, Y.H., and Beardmore, D.H. 1980. Major chemical and physical features of cellulosic materials as substrates for enzymatic hydrolysis. *Adv. Biochem. Eng.*, 14:101-117.
- [20] Mandels, M., Hontz, L., and Nystrom, J. 1974. Enzymatic hydrolysis of waste cellulose. *Biotechnol. Bioeng.*, 16:1471-1493.
- [21] Millet, B.A., Baker, A.J., and Slater, L.D. 1975. Pretreatment to enhance chemical, enzymic and microbiological attack on cellulosic materials. *Biotechnol. Bioeng. Symp.*, 5:193-219.
- [22] Dunlap, C.E., Thomson, J., and Chang, L.C. 1976. Treatment processes to increase cellulose digestibility. *AIChE Symp. Ser.*, 72(158):58-63.
- [23] Tussanari, T. and Macey, C. 1977. Differential two speed roll mill pretreatment of cellulosic materials for enzymatic hydrolysis. *Biotechnol. Bioeng.*, 19:1321-1330.
- [24] Chang, H.M., Chang, Y.C., and Tsao, G.T. 1981. Structure, pretreatment and hydrolysis of cellulose. *Adv. Biochem. Eng.*, 20:15-42.
- [25] McBurney, L.F. 1954. Kinetics of degradation reactions. In *High polymers, vol. 5, Pt. 1. Cellulose and cellulose derivatives*, pp. 99-129. Ott, E., Spurlin, H.M. and Graffin, M.W., Eds. Interscience Publishers, New York.
- [26] Wannicker, J.O., Jeffries, R., Colbran, R.L., and Robinson, R.N. 1966. A Review of the literature on the effect of caustic soda and other swelling agents on the fine structure of cotton. *Shirley Inst. Pamphlet. No.93*, 1979. Shirley Inst. Didsbury, Manchester.
- [27] Price, E.C. and Cheremisinoff, P.N. 1981. Biogas production and utilization. *Ann. Arbor. Science*, 47.
- [28] Callihan, C.D. 1970. How engineers are putting microbes to work. *Chemical Engineering*, 77(20):161.
- [29] Han, Y.W., Dunlap, C.E., and Callihan, C.D. 1971. Single cell protein from cellulose wastes. *Food Tech.*, 25:130-134, 154.
- [30] Srinivasan, V.R. and Han, Y.W. 1969. Utilization of bagasse. In *Cellulases and their applications. Adv. chem. ser.*, vol. 95, pp. 447-460. Gould, R.F., Ed. American Chem. Soc., Washington, D.C.
- [31] Callihan, C.D. and Dunlap, C.E. 1969. The economics of protein production from cellulosic wastes. *Compost Science*, 10:1-2.
- [32] Callihan, C.D. and Dunlap, C.E. 1971. Pilot production of SCPs from cellulosic wastes. *Report No. SW-24C*. US Environmental Protection Agency. Supdt. of Doc. Stock No. 5502-0027.
- [33] Han, Y.W. 1969. Studies of the bacterial enzymes involved in the degradation of cellulose. Ph.D. Dissertation. Louisiana State University.
- [34] Han, Y.W. and Srinivasan, V.R. 1968. Isolation and characterization of a cellulose utilizing bacterium. *Appl. Microbiol.*, 16(8):1140-1145.
- [35] Selby, K. and Maitland, C.C. 1967. The cellulase of *Trichoderma viride*. Separation of the components involved in the solubilization of cotton. *Biochem J.*, 104-716.
- [36] Wood, T.M. 1968. Cellulolytic enzyme system of *Trichoderma koningii*. *Biochem. J.*, 109:217.
- [37] Wood, T.M. 1969. The cellulase of *Fusarium solani*. Resolution of the enzyme complex. *Biochem. J.*, 115:457.
- [38] Wood, T.M. 1972. *Proc. IV IFS. Ferment. Technol. Today*, p. 711. Society of Fermentation Technology, Japan.
- [39] Okazaki, M. and Moo-Young, M. 1978. Kinetics of enzymatic hydrolysis of cellulose: Analytical description of mechanistic model. *Biotechnol. Bioeng.*, 20:637-663.
- [40] Reese, E.T. 1959. *Marine boring and fouling organisms*. University of Washington Press, Seattle. 265.
- [41] Tsao, G.T., Ladisch, M., Ladisch, C., Hsu, T.A., Dale, B., and Chou, T. 1978. Fermentation substrates from cellulosic materials: Production of fermentable sugars from cellulosic materials. *Ann. Reports of Fermentation Processes*, 2:1-22.
- [42] Desai, A.J. and Betrabet, S.M. 1972. Cellulase activity of microorganisms isolated from cotton deteriorated during storage. *Indian Journal of Biochemistry and Biophysics*, 9:212-214.
- [43] Banik, B., Mishra, A.K., and Nanda, G. 1989. Partial purification and properties of extracellular cellulase from *Asp. fumigatus*. *Transactions of Bose Research Institute (India)*, 52:43-55.
- [44] Suzuki, H., Yamane, K., and Nisizuwa, K. 1969. In *Cellulases and their applications, Advances in Chem. Series*, vol. 95, p. 60. Gould, R.F., Ed. American Chemical Society Pub., Washington, D.C.
- [45] Suzuki, H. 1975. Cellulose formation in *Pseudomonas fluorescens var. cellulosa*. In *Symposium on enzymatic hydrolysis of cellulose*, pp. 151-169. Bailey, M., Enari, T.-M., and Linko, M., Eds. SITRA, Aulanko, Finland.
- [46] Simmons, E.G. 1977. *Proceedings of the Second International Mycological Congress*, p. 618. Tampa, Florida.
- [47] Halliwell, G. and Griffin, M. 1973. The nature and mode of action of the cellulolytic component C₁ of the *Trichoderma koningii* on native cellulose. *Biochemical J.*, 135:587.
- [48] Halliwell, G. 1965a. Catalytic decomposition of cellulose under biological conditions. *Biochemical J.*, 95:35.
- [49] Halliwell, G. 1965b. Hydrolysis of fibrous cotton and reprecipitated cellulose by cellulolytic enzymes from soil microorganisms. *Biochemical J.*, 95:270.
- [50] Wood, T.M. and McCrae, S.I. 1972. The purification and properties of the C₁ component of *Trichoderma koningii* cellulase. *Biochem. J.*, 128:1183.
- [51] Wood, T.M. and McCrae, S.I. 1975a. In *Symposium on enzymatic hydrolysis of cellulose*, pp. 231-254. Bailey, M., Enari, T.-M. and Linko, M., Eds. SITRA, Aulanko, Finland.
- [52] Wood, T.M. and McCrae, S.I. 1978. The cellulase of *Trichoderma koningii*. *Biochemical J.*, 171:61.
- [53] Selby, K. 1968. In *Biodeterioration of materials*, p. 62. Walters, A.H. and Elphick, J.J., Eds. Elsevier Publishing Co., Amsterdam-London-New York.
- [54] Wood, T.M. and McCrae, S.I. 1977b. In *Proceedings of Bioconversion Symposium*, p. 111. Ghose, T.K., Ed. IIT, Hauz Khas, New Delhi-29, India.
- [55] Wood, T.M. 1971. The cellulase of *Fusarium solani*. *Biochem. J.*, 121:353-362.
- [56] Wood, T.M. and McCrae, S.I. 1977a. *Carbohydrate Research*, 57:117-133.
- [57] Marsden, W.L. 1986. *CRC Critic. Rev. in Biotechnol.*, vol. 3, issue 3, CRC Press, Boca Raton, Florida.
- [58] Donefer, E., Adeleye, I., and Jones, T. 1969. Effect of urea supplementation on the nutritive value of NaOH-treated oat straw. In *Adv. Chem. Ser.*,

- vol. 95, p. 328-342. Gould, R.F., Ed. ACS Publications, Washington, D.C.
- [59] Southgate, D.A.T. 1976. *Determination of food carbohydrates*, pp. 91-93. Applied Science, London.
- [60] Zahorsky O.R. 1981. *CRC handbook of biosolar resources*, vol. II. CRC Press, Boca Raton, Florida.
- [61] Chahal, D.S. and Gray, W.D. 1968. In *Biodeterioration of materials-microbiological and allied aspects*, pp. 584-593. Walter, A.H. and Elphic, J.S., Eds. Elsevier Publ. Co., Barking Essex, England.
- [62] Chang, S.T., Khor, G.L., Ng, C.L., Ong, K.C., Quimio, T.H., Stanton, W.R., and Wang, W.C.-W. 1983. Mushrooms: Producing single cell protein on lignocellulosic or other food and agricultural wastes. In *Handbook of indigenous fermented foods*, pp. 573-604. Steinkraus, K.H., Ed. Marcel Dekker, New York.
- [63] Enriques, A. and Rodrigues, M. 1983. High productivity and good nutritive value of cellulolytic bacteria grown on sugarcane bagasse. *Biotech. Bioeng.*, 25:877-880.
- [64] Rodrigues, H., Alvarez, R., and Enriques, A. 1993. Evaluation of different alkali treatments of bagasse pith for cultivation of *Cellulomonas* sp. *World J. of Microbiol. and Biochem.* 9(2):213-215.
- [65] Azzam, A.M., Ghoneim, S., and Ebrahim, M.Z. 1990. Pretreatments of agro-cellulosic waste (bagasse) for microbial biomass production with a defined mixed culture (*T. viride* & *C. utilis*). *Food Biotechnol.* 4(1):474.
- [66] Azizi, K.A., Shamsala, T.R., and Sreekantiah, K.R. 1990. Cultivation of *Pleurotus sajor-caju* on certain agro-industrial wastes and utilisation of the residues for cellulose and D-xylanase production. *Mushroom J. for the Tropics*, 10(1):21-26.
- [67] Callihan, C.D. and Clemmer, J.E. 1979. Biomass from cellulosic materials. In *Microbial biomass-economic microbiology*, vol. 4, chap. 9, pp. 271-288. Rose, A.H., Ed. Academic Press, New York.
- [68] Crawford, D.L., McCoy, E., Harkin, J.M., and Jones, P. 1973. Microbial protein from waste cellulose by *Thermomonospora fuscus*. A thermophilic actinomycete. *Biotechnol. Bioeng.*, 15:833-843.
- [69] Elisashvili, V.I., Glonti, N.M., Kachilishvili, E.T., Kiknodze, M.O., and Tusishvili, K.A. 1992. Selection of higher basidiomycetes—Protein and enzyme procedures. *Appl. Biochem. and Microbiol.*, 28(3):271-274.
- [70] Kokhzeidre, N.G. and Elisashvili, V.I. 1993. Lignocellulolytic activity of *Pleurotus ostreatus* IBK 191 in the solid phase fermentation of the waste of tea production. *Appl. Biochem. and Microbiol.*, 29(2):169-173.
- [71] Labeneish, M.E.O., Abou-Donia, S.A., Mohamed, M.S., and El Zlaki, E.M., 1960. *Expt. Stat. Bull.* 622.
- [72] Nancib, N., Nancib, A., and Boudrant, J. 1997. Use of waste date products in the fermentative formation of baker's yeast biomass by *S. cerevisiae*. *Bioresource Technology*, 60(1):67-71.
- [73] Sturion, G.L. and Oetterer, M. 1995. Utilization of banana leaves as substrate on edible mushroom cultivation (*Pleurotus* spp.). *Ciencia e Tecnologia de Alimentos*, 15(2):194-200.
- [74] Burwell, J.A. and Shu-Ting Chang. 1994. Biomass and extracellular hydrolytic enzyme production by six mushroom species grown on soybean waste. *Biotechnology Letters*, 12:1317-1322.
- [75] Nazarenko, A.V., Sokolov, V.N., Ginak, A.I., and Ostrer, B.S. 1994. Biosynthesis of protein and enzymes of the cellulolytic complex by microfungus *Asp. sp.* on comcob. *Appl. Biochem. and Microbiol.*, 29(3):331-334.
- [76] Umar Duhot, M., Yakoub Khan, M., and Yousuf Khan, M. 1994. Effect of temperature and pH on the production of single cell protein by *Penicillium expansum*. *Science International*, 5(3):259-261.
- [77] Choi, M.H. and Park, Y.H. 1998. The influence of yeast thermophilic composting of food waste. *Letters in Applied Microbiology*, 26:175-178.
- [78] Nakasaki, K., Yaguchi, H., Sasaki, Y., and Kubota, H. 1990. Effect of oxygen concentration on composting of garbage. *J. of Fermentation and Bioeng.*, 70:431-433.
- [79] Nakasaki, K., Yaguchi, H., Sasaki, Y., and Kubota, H. 1992. Effect of C/N ratio on thermophilic composting of garbage. *J. of Fermentation and Bioeng.*, 73:43-45.
- [80] Hussein, A.M., El Saied, H., and Yasin, M.H. 1992. Bioconversion of hemicelluloses of rice hull black liquor into single cell protein. *J. of Chemical Technol. and Biotechnol.*, 53(2):147-152.
- [81] Bellarmy, W.D. 1969. Cellulose as a source of single-cell-proteins—A preliminary evaluation. *General Electric Research and Development Center Report No. 69-C-335*. Schenectady, General Electric Company, 5.
- [82] Paminent, N., Robinson, C.W., Hilton, J., and Moo-Young, M. 1978. Solid-state cultivation of *Chaetomium cellulolyticum* on alkali-pretreated sawdust. *Biotechnol. Bioeng.*, 20:1735-1744.
- [83] Palmer, F.G. 1976a. The feeding value of straw to ruminants. *ADASQ Rev.*, 21:220-234.
- [84] Palmer, F.G. 1976b. The feeding value and worthwhileness of chemically processed straw for ruminants. *ADASQ Rev.*, 22:247-266.
- [85] Srinivasan, V.R. and Callihan, C.D. 14 December 1971. US Patent 3 627 095.
- [86] Dyer, J.A., Riquelme, E., Baribo, L., and Couch, B.Y. 1975. Waste cellulose as an energy source for animal protein production. *World Animal Review*, 15:39-43.
- [87] Hesseltrine, C.W. 1972. Solid state fermentation. *Biotechnol. Bioeng.*, 14:517-532.
- [88] Brown, D.E. and Fitzpatrick, S.W. 1976. Food from waste paper. In *Food from waste*, chap. 9, pp. 139-155. Birch, G.G., Parker, K.J., and Worgan, J., Eds. Applied Science, New York.
- [89] Kamikubo, T., Tanaka, M., Taniguchi, M., Morita, T., and Matsumo, R. 1980. Production of SCP from cellulose. In *Adv. in Biotechnol.*, vol. 2, pp. 311-318. Moo-Young, M. and Robinson, C.W., Eds. Pergamon Press, Canada.
- [90] Kuzantskheiyi, M.S., Zvyagil'skaya, R.A., Rainina, E.I., and Robinovich, M.I. 1991. Enzymatic hydrolysis of cellulosic wastes of tea concentrate production and use of hydrolyzates to produce microbial protein. *Applied Biochemistry and Microbiology*, 27(5):518-523.
- [91] Dahot, M.U. and Ahro, A.Q. 1994. Biosynthesis of lysine and histidine by *Penicillium expansum* using agricultural waste as a carbon source. *Science International*, 6(1):63-66.
- [92] Moo-Young, M. 1976. A survey of SCP production facilities. *Process Biochem.*, 11:10, 32.
- [93] Skogman, H. 1976. In *Food from waste*. Birch, G.G., Parker, K.J., and Worgan, T.G., Eds. Appl. Sci. Publ. London.
- [94] Wu, Y.V. 1989. Protein rich residue from ethanolic fermentation of high lysine, dent, waxy and white corn varieties. *Cereal Chemistry*, 66(6):506-509.
- [95] Callihan, C.E. and Srinivasan, V.R. 1973. *Final Report on Grant FPO 03284 to the Federal Solid Waste Management Programmes*, US Environmental Protection Agency.
- [96] Molina, O.E., Peranti de Galvez, N.I., Frigerio, C.I., and Cordoba, R.R. 1984. Single cell protein production from bagasse pith pretreated with NaOH at room temp. *Appl. Microbiol. and Biotechnol.*, 20:335-339.
- [97] Lee, S.M. and Kim, Z.U. 1992. Extraction of proteins from soy milk residue using enzymes from *Asp. oryzae*. *J. of the Korean Agri. Chem. Soc.*, 35(1):64-67.
- [98] Trifonova, V.V., Ignatova, N.I., Milyukova, T.B., Overchenko, M.B., and Rimareva, L.V. 1993. Possibility of utilizing various types of fruit and vegetable raw material for microbial synthesis of lysine. *Appl. Biochem. and Microbiol.*, 29(4):429-432.
- [99] Garg, N., Tandon, D.K., and Kalra, S.K. 2000. Protein enrichment of mango peel through solid state fermentation using *Aspergillus niger* for utilization as feed. *Indian Food Packer*, 54(3):62-64.
- [100] Sethi, R.P. 1978. The conversion of reject banana and mango stone into animal feed using solid substrate fermentation. Ph.D. Thesis. Dept. of Microbiology, Punjab Agri. University, Ludhiana, India.
- [101] Davy, C.A.E., C. Eng., M.I., and Chem. E. 1981. Recovery of fruit and vegetable waste. In *Food industry wastes: Disposal and recovery*, pp. 219-230. Herzka, A. and Booth, R.G., Eds. Applied Science Publishers.
- [102] Bhalla, T.C. and Joshi, M. 1994. Protein enrichment of apple pomace by co-cultures of cellulolytic moulds and yeasts. *World J. of Microbiol. and Biotechnol.*, 10(1):116-117.

- [103] Illanes, A., Aroca, G., Cobello, L., and Acevedo, F. 1992. Solid substrate fermentation of leached beet pulp with *Trichoderma aureoviride*. *World J. of Molecular Biol. and Biotechnol.*, 8(5):488-493.
- [104] Kuzmanova, S., Vandeska, E., and Dimitrovski, A. 1991. Production of mycelial protein and cellulolytic enzymes from food wastes. *J. of Industr. Microbiol.*, 7(4):257-261.
- [105] Grewal, H.S., Kalra, K.L., and Kahlon, S.S. 1990. Citrus (Kinnow-Mandarin) residue as potential substrate for single cell protein. *J. of Research*, 27(1):90-96.
- [106] Raimbault, M. and Alazard, D. 1980. Culture method of study fungal growth in solid state fermentation. *Evr. J. Appl. Microbiol. Technol.*, 9:199-205.
- [107] Kunhi, A.A.M., Ghildyal, N.P., Lonsaie, B.K., Ahmed, S.Y., and Natarajan, C.P. 1981. Studies on the production of alcohol from saccharified waste residues from cassava starch processing industries. *Die Starke*, 33(8, 5):275-279.
- [108] Limes, K.K. 1972. *U.S. Patent* 4-144, 132-139.
- [109] Rolz, C. 1975. In *Single cell protein II*, pp. 273-313. Tannenbaum, S.R. and Wang, D.I.C., Eds. MIT Press, Cambridge, Massachusetts.
- [110] de Cabrera, S., Mayorga, H., Espinosa, R., and Rolz, C. 1976. *Proc. Int. Congr. Food. Sci. Technol.* 2974, vol. 4, pp. 296-301.
- [111] Selim, M.H., Elshafei, A.M., and El-Diwany, A.I. 1991. Production of single cell protein from yeast strains grown in Egyptian vinasse. *Bioresource Technology*, 36(2):157-160.
- [112] Krishna, C. and Chandrasekaran, M. 1995. Economic utilization of cabbage wastes through solid-state fermentation by native microflora. *J. of Food Sc. and Tech. (India)*, 32(3):199-201.
- [113] Kristensen, T.P. 1978. Continuous SCP production from *Cellulomonas* sp. and *Candida utilis* grown in mixture on barley straw. *Euro. J. Appl. Microbiol.*, 5:155-163.
- [114] Chang, W.T.M., Hsu, W.-H., Lai, M.N., and Chang, P.P. 1980. Production of SCP from rice hulls for animal feed. *Dev. Ind. Microbiol.*, 21:313-325.
- [115] Chahal, D.S., Vlach, D., and Moo-Young, M. 1981. Upgrading the protein feed value of lignocellulosic material using *Chaetomium cellulolyticum* in solid-state fermentation. In *Advances in Biotechnol.*, vol.-II, pp. 327-332. Moo-Young, M. and Robinson, C.W., Eds. Pergamon Press, Canada.
- [116] Khor, G.L., Alexander, J.C., Santos-Nuncz, J., Reade, A.E., and Gregory, K.F. 1976. Nutritive value of thermotolerant fungi grown on cassava. *J. Inst. Can. Sci., Technol. Aliment.*, 9:139-143.
- [117] Reade, A.E. and Gregory, K.F. 1975. High temperature production of protein enriched feed from cassava by fungi. *Appl. Microbiol.*, 30:897-904.
- [118] Moo-Young, M., Daugulis, A.G., Chahal, D.S., and MacDonald, D.G. 1979. The Waterloo process for SCP production from waste biomass. *Process Biochem.*, 14(10):38-40.
- [119] Macris, B.J. and Kokke, R. 1978. Continuous fermentation to produce fungal protein. Effect of growth rate on the biomass yield and chemical composition of *Fusarium moniliforme*. *Biotechnol. Bioeng.*, 20:1027-1035.
- [120] Gregory, K.F., Reade, A.E., Santos-Nunez, Alexander, J.C., Smith, R.E., and McClean, J. 1978. Further thermotolerant fungi for the conversion of Cassava starch to protein. *Annual Feed Sci. Tech.*, 2:7-19.
- [121] Han, Y.W. and Anderson, A.W. 1975. Semisolid fermentation of ryegrass straw. *Appl. Microbiol.*, 30:930-934.
- [122] Worgan J.T. 1976. Wastes from crop plants as raw materials for conversion by fungi to food or livestock feed. In *Food from wastes*, pp. 23-41. Birch, G.G., Parker, K.T. and Worgan, J.T., Eds. Appl. Publishers, London.
- [123] Srinivasan, V.R. 1975. Production of bioprotein from cellulose. In *Sym. on enzymatic hydrolysis of cellulose*, pp. 81-109. Bailey, M., Enari, T.-M., and Linko, M., Eds. The Finnish National Fund for R&D(SITRA), Helsinki.
- [124] Shoemaker, S., Watt, K., Tristovsky, G., and Cox, R.V. 1983a. Characterization and properties of cellulases purified from *Trichoderma reesei* strain L27. *Biotechnol.*, 1:687.
- [125] Tuse, D., Russell, L.A., and Hsich, D.F.H. 1981. Nutritional and toxicological evaluation of SCP produced from an environmental waste. In *Advances in Biotechnology*, vol. II, pp. 363-368. Moo-Young, M. and Robinson, C.W., Eds. Pergamon Press, Canada.
- [126] Sifton, O.C., Magruder, G.C., Book, N.L., and Graddy, J.L. 1980. Comparison of immobilized cell reactor and CSTR for ethanol production. *Biotech. Bioeng. Symp.*, 10:213-239.
- [127] Fan, L.T., Gharpurey, M.M., and Lee, Y.H. 1981. Evaluation of pretreatments for enzymatic conversion of agricultural residues. *Biotech. Bioeng. Symp.*, 11:29-45.
- [128] Mandels, M., Dorval, S., and Medeiros, J. 1978. Saccharification of cellulose with *Trichoderma cellulase*. In *Proc. of the second fuels from biomass symp.*, p. 627. Wiley, New York.
- [129] Sanchez, P., Rigol, L., Gaset, A., and Mailhol, P. 1992. Process and device for preparation of pentose and/or hexose syrup from plant materials rich in hemicellulose. *French Patent Application FR. 2668 165 A.1.*
- [130] Rao, J.S., Singhal, R.S., and Kulkarni, P.R. 1998. Xylose syrup from cottonseed hulls-optimization of hydrolysis conditions and purifications by ion exchange resins. *J. of Scient. and Indust. Res.(I)*, 57:196-200.
- [131] Pellerin, P., Gosselin, M., Lepoutre, J.P., Samain, E., and Debetre, P. 1991. Enzyme production of oligosaccharides from corn cob xylan. *Enzyme and Microbial Technology*, 13(8):617-621.
- [132] Kvesitadze, E.G., Adeishvili, E.T., and Tkeshelashvili, R.Sh. 1995. Factors influencing the biosynthesis of thermostable endoglucanases and xylanase in a culture of the fungus *Allesteria terrestris*. *Appl. Biochem. and Microbiol.*, 30(4,5):476-479.
- [133] Toyama, N. and Ogawa, K. 1975. Sugar production from agricultural woody wastes by saccharification with *T. viride* cellulase. *Biotechnol. Bioeng. Symp.*, 5:225-244.
- [134] Primo-Yufera, E., Gil-Tortosa, C.I., and Garcia-Breijo, F.J. 1995. Hydrolysis of corn cob lignocellulosic residue for pentose preparation. *Bioresource Tech.*, 52(1):1-4.
- [135] Okeke, B.C. and Obi, S.K.C. 1995. Saccharification of agrowaste materials by fungal cellulases and hemicellulases. *Bioresource Tech.*, 51(1):23-27.
- [136] Leathers, T.D. and Gupta, S.C. 1996. Saccharification of corn fibre using enzymes from *Aureobasidium* sp. strain NRRL Y-2311-1. *Appl. Biochem. and Biotech.*, 59(3):337-347.
- [137] Moszozynski, P. and Pyc, R. 1991. Enzymatic saccharification of the solid waste fraction from extraction of coffee beans. *Przemysl Spozycwcy*, 45(5,6):146-149.
- [138] Beerh, O.P., Raghuramaiah, Krishnamurthy, G.V., and Giridharilal, N. 1976. Utilization of mango waste: Recovery of juice from waste pulp and peel. *J. Food. Sci. and Technol. (India)*, 13(3):138-141.
- [139] Sutherland, I.W. 1996. Microbial biopolymers from agril products: Production and potential. *International J. Biodeterioration and Biodegradation*, 38(3,4):249-261.
- [140] Israelides, C., Scanlon, B., Smith, A., Harding, S.E., and Jumel, K. 1994. Characterization of pullulans produced from agro-industrial wastes. *Carbohydrate Polymers*, 25(3):203-209.
- [141] Roukas, T. and Billaderis, C.G. 1995. Evaluation of carob-pod as a substrate for pullulan production by *Aureobasidium pullulans*. *Appl. Biochem. and Biotech.*, 55(1):27-44.
- [142] Bilanovic, D., Shelef, G., and Green, M. 1994. Xanthan fermentation of citrus waste. *Biores. Technol.*, 48(2):169-172.
- [143] Green, M., Sheleb, G., and Bilanovic, D. 1994. Effect of various citrus waste fraction on xanthan fermentation. *Chemical Engg. J.*, 56(1):B37-B41.
- [144] Renard, C.M.G.C., Lemeunier, C., and Thibault, J.F. 1995. Alkaline extraction of xyloglucan from depectinised apple pomace: Optimization and characterisation. *Carbohydrate Polymers*, 28(3):209-216.
- [145] Abetyan, V.A., Chu, D.C., and Yamamoto, T. 1995. Production of cyclodextrin from rice bran. *Applied Biochem. and Microbio.*, 31(4):387-390.
- [146] Sanderson, G.R. and Clark, R.C. 1983. Laboratory-Produced microbial polysaccharide has many potential food applications as a gelling, stabilizing, and texturizing agent. *Food Technol.*, 37:63-70.
- [147] Baird, J.K., Sandford, P.A., and Cottrell, I.W. 1983. Industrial applications of some new microbial polysaccharides. *Biotechnology*, 1:778-783.

- [148] Woodbine, M. 1959. Microbial fat: Microorganisms as potential fat producers. *Prog. in Industrial Microbiology*, 1:181.
- [149] Ratledge, C., Boulton, C.A., and Evans, C.T. 1984. In *Continuous cultures of microorganisms*, vol. 8, pp. 272-291. Dean, A.C.R., Ellwood, D.C., and Evans, C.G.T., Eds. Ellis Horwood, Chichester.
- [150] Boulton, C.A. and Ratledge, C. 1981. Correlation of lipid accumulation in yeasts with possession of ATP: Citrate lyase. *J. Gen. Microbiol.*, 127:169-176.
- [151] McLee, A.G., Kornendy, A.C., and Wayman, M. 1972. Isolation and characterization of n-butanol-utilizing microorganisms. *Can. J. Microbiol.*, 18:1191-1195.
- [152] Wayman, M., Jenkins A.D., and Kornendy, A.C. 1984. In *Biotechnology for the oils and fats industry*, pp. 129-143. Ratledge, C., Rattray, J.B.M., and Dawan, P.S.S., Eds. Monograph no. 1. Am. Oil Chem. Soc.
- [153] Eydokimava, G.A., Raisine, G.I., Kustykevich, L.I., Lyakh, V.V., Zalashko, M.V., Gurinovichi, E.S., Bogdanovskaya, Zh.N., and Obraztsova, N.V. 1974. *Prikl. Biokhim. Mikrobiol.*, 10:780.
- [154] Andreyevskaya, V.D. and Zalashko, M.V. 1979. *Gidroliz Lesokhim. Prom-st.*, 8:3.
- [155] Gogolewski, M., Nogala-Kalucka, M., and Makarewicz, P. 1996a. Biosynthesis of lipids and their changes during culture of selected yeast strains on carrot juice I. *J. of Japan Oil Chem. Soc.*, 45(6):555-560.
- [156] Gogolewski, M., Nogala-Kalucka, M., Makarewicz, P., and Warowicz, E. 1996b. Biosynthesis of lipids and their changes during culture of selected yeast strains on carrot juice II. *J. Jap. Oil Chem. Soc.*, 45(6):561-567.
- [157] Gomes, T. and Caponio, F. 1997. Evaluation of the state of oxidation of crude olive-pomace oils. Influence of olive-pomace drying and oil extraction with solvent. *J. Agril. and Food Chem.*, 45(4):1381-1384.
- [158] Kennedy, M.J. 1994. Applied pomace and kiwifruit: processing options (Rev.). *Austr. Biotechnol.*, 4(1):43-49.
- [159] McNary, R.R. and Dougherty, M.H. 1960. Citrus vinegar. *Florida Univ. Agril. Expt. Stat. Bull.*, 622.
- [160] Kumar P.K.R., Singh, A., and Schugerl, K. 1991. Fed-batch culture for the direct conversion of cellulosic substrates of acetic acid/ethanol by *Fusarium oxysporum*. *Process Biochem.*, 26(4):209-216.
- [161] Spurgin, M.M. 1964. Vinegar base production from waste pineapple juice. *Queens Land J. Agril. Sci.*, 21:213.
- [162] Richardson, K.C. 1967. Submerged acetic acid production of a vinegar base produced from waste pineapple juice. *Biotech. and Bioeng.*, 9:171.
- [163] Saddler, J.N. and Khan, A.W. 1979. Cellulose degradation by a new isolate from sewage sludge, a member of the *Bacteroidaceae* family. *Canadian J. of Microbiol.*, 25:1427-1431.
- [164] Manonmani, H.K. and Sreekantiah, K.R. 1987. Studies on conversion of cellulose hydrolysate into citric acid by *Asp. niger*. *Process Biochem.*, 22:92-94.
- [165] Menezes, T.J.B., Salva, T.J.B., Baldini, V.L., Papini, R.S., and Sales, A.M. 1989. Protein enrichment of citrus waste by solid-state fermentation. *Process Biochem.*, 24:167-171.
- [166] Tran, C.T. and Mitchell, D.A. 1995. Pineapple waste—A novel substrate for citric acid production by solid-state fermentation. *Biotechnol. Letters*, 17(10):1107-1110.
- [167] Hang, Y.D. and Woodams, E.E. 1984. Apple pomace: A potential substrate for citric acid production by *Asp. niger*. *Biotechnol. Letters*, 6:763-764.
- [168] Hang, Y.D. and Woodams, E.E. 1986a. Solid state fermentation of apple pomace for citric acid production. *MIRCEN. J. Appl. Microbiol. Biotechnol.*, 2:283-287.
- [169] Hang, Y.D. and Woodams, E.E. 1986b. Utilization of grape pomace for citric acid production by solid-state fermentation. *Am. J. Enol. Vitic.*, 37:141-142.
- [170] Hang, Y.D. and Woodams, E.E. 1987. Microbial production of citric acid by solid-state fermentation of kiwi fruit peel. *J. Food Sci.*, 52:226-227.
- [171] Garg, N., Tandon, D.K., and Kalra, S.K. 1998. Potential of using guava jelly processing waste for citric acid production through microbial fermentation. *Indian Food Packer (India)*, 523:7-9.
- [172] Thomopoulos, C.D. 1994. Fermentation of orange processing wastes for citric acid production. *J. of Sc. of Food and Agric.*, 65(1):117-120.
- [173] Shankaranand, V.S. and Lonsane, B.K. 1994. Coffee husk: An inexpensive substrate for production of citric acid by *Aspergillus niger* in a SSF system. *World J. of Molecular Biol. and Biotechnol.*, 10(2):165-168.
- [174] Garg, N. and Hang, Y.D. 1995. Microbial production of organic acids from carrot processing waste. *J. of Food Sc. and Tech. (India)*, 32(2):119-121.
- [175] El-Sharkawy, S.H., Karim, M.I.A., and Wong Su Yin. 1995. Production of citric acid from cocoa juice waste. *ASEAN Food Journal*, 10(3):112-114.
- [176] Xavier, S. and Lonsane, B.K. 1994. Sugar cane press mud as a novel and inexpensive substrate for production of lactic acid in a solid-state fermentation system. *Appl. Microbiol. & Biotechnol.*, 41(3):291-295.
- [177] Schopmeyer, H.H. 1954. Lactic acid. In *Industrial fermentations*, pp. 391-419. Underkofler, L.A. and Hickey, R.J., Eds. Chemical Publishing Co., New York.
- [178] Leonard, R.H., Peterson, W.H., and Johnson, M.J. 1948. Lactic acid from fermentation of sulfite waste liquor. *Ind. Eng. Chem.*, 40:57-67.
- [179] Pekar, I. 1994. The obtaining of tartaric acid from grape pomace and determination of tannin. *Gida*, 19(1):23-25.
- [180] Carta, F.S., Socoli, C.R., Machado, L., and Machado, C.M.M. 1990. Prospect of using cassava bagasse waste for producing fumaric acid. *J. of Scient. and Industr. Res.*, 57:644-649.
- [181] Clausen, E.C. and Gaddy, J.L. 1981. Fermentation of biomass into acetic and propionic acids with *Propionibacterium aceti-propionii*. In *Advan. in Biotechnol.*, vol. II, pp. 63-69. Moo-Young, M. and Robinson, C.W. Eds., Pergamon, Toronto.
- [182] Playne, M.J. 1981. Volatile fatty acid production by anaerobic fermentation of lignocellulosic substrates. In *Advan. Biotechnol.*, vol. II, pp. 85-90. Moo-Young, M. and Robinson, C.W., Eds. Pergamon, Toronto.
- [183] Sanderson, J.E., Wise, D.L., and Augenstein, D.C. 1979. Organic chemicals and liquid, fuels from algal biomass. *Biotechnol. Bioeng. Symp.*, 8:131-151.
- [184] Tomlin, D.C., Johnson, R.R., and Dehority, B.A. 1965. Relationship of lignification to in-vitro cellulose digestibility of grasses and legumes. *J. Ani. Sc.*, 24:161.
- [185] Feist, W.C., Baker, A.J., and Turkow, H. 1970. Alkali requirements for improving digestibility of hardwoods by rumen microorganisms. *J. Ani. Sc.*, 30:832.
- [186] Cross, H.H., Smith, L.W., and Debnath, J.V. 1974. Rates of in-vitro forage fiber digestion as influenced by chemical treatment. *J. Ani. Sc.*, 39:4.
- [187] Han, Y.W., Lee, J.S., and Anderson, A.W. 1975b. Chemical composition and digestibility of rye grass straw. *J. Agri. Food Chem.*, 23:928-931.
- [188] Baumgardt, B.R., Taylor, M.W., and Cason, J.L. 1962. Evaluation of forages in the laboratory. II. Simplified artificial rumen procedure for obtaining repeatable estimates of forage nutritive value. *J. Dairy Sc.*, 45(1):62-68.
- [189] Guggolz, J.R., Saundress, R.M., Kohler, G.O., and Klopfenstein, T. 1971b. Enzymatic evaluation of process for improving agricultural wastes for ruminant feeds. *J. Ani. Sc.*, 33:167-170.
- [190] Anderson, D.C. and Ralston, A.T. 1973. Chemical treatment of rye grass straw: in-vitro dry matter digestibility and compositional changes. *J. Ani. Sc.*, 37:148-152.
- [191] Graham, R.P. 1972. *U.S. Patent 3:692, 530*.
- [192] Millet, B.A., Baker, A.J., Feist, W.C., Meilenberger, R.W., and Slatter, L.D. 1970. Modifying wood to increase its in-vitro digestibility. *J. Ani. Sc.*, 31:781.
- [193] Turkow, H. and Feist, W.C. 1969. A mechanism for improving the digestibility of lignocellulosic materials with dil. alkali and liq. ammonia. In *Adv. in chem. ser. no. 95*, pp. 197-218. Amer. Chem. Soc. Pub., Washington.
- [194] Guggolz, J., Kohler, G.O., and Klopfenstein, T. 1971a. Composition and improvement of grass straw for ruminant nutrition. *J. Ani. Sc.*, 33(1):151-156.
- [195] Weiss, A.C., Jr., Guggolz, J., Kohler, G.O., Walker, H.G. Jr., and Currett, W.N. 1972. Improving digestibility of straws for ruminant feed by aqueous ammonia. *J. Ani. Sc.*, 35(1):109-112.

- [196] Matsuo, S. 3 August 1971. U.S. Patent 3 597 218.
- [197] Barrows, L., Seal, K.J., and Eggins, H.D.W. 1979. Biodegradation of barley straw by *Coprinus cinereus* for the production of ruminant feed. In *Straw decay and its effect on utilization and disposal*, p. 147. Grassbald, F., Ed. John Wiley & Sons Ltd., Sussex, UK.
- [198] Stone, J.E., Scullian, A.M., Dunfer, E., and Ahlgren, E. 1969. Digestibility as a simple function of a molecule of similar size to a cellulase enzyme. *Adv. in Chem. Ser.*, 95:219.
- [199] Han, Y.W. and Callihan, C.D. 1974. Cellulose fermentation: Effect of substrate pretreatment on microbial growth. *Appl. Microbiol.*, 27:159.
- [200] Han, Y.W., Pence, J.W., and Anderson, A.W. 1975a. U.S. Patent 3:937 849, 1976a.
- [201] Bishop, R.W. 22 June 1971. U.S. Patent 3, 586 511.
- [202] Miller, R.B. and Laurie, C.K. 20 March 1973. U.S. Patent 3 721 567.
- [203] Worden, W.W. 17 August 1971. U.S. Patent 3 600 190.
- [204] Tadeu-Pontes, M.A., Carvalheiro, F., Roseiro, J.C., and Amaral-collaco, M.T. 1996. Evaluation of product composition profile during an extrusion based process of tomato pomace transformation. *Agro Food Industry Hi-tech*, 7(3):39-40.
- [205] Rahmat, H., Hodge, R.A., Manderson, G.J., and Yu, P.L. 1995. Solid substrate fermentation of *Kloeckera apiculata* and *Candida utilis* on apple pomace to produce an improved stock-feed. *World J. of Microbiol. and Biotechnol.*, 11(2):168-170.
- [206] Henn, T. and Kunz, B. 1996. Use of plant pomace for manufacture of functional drinks. *Flussiges Obst*, 63(12):715-719.
- [207] Takamami, S., Ohsawa, K., Kuwabara, H., and Kurokouchi, K. 1996. Studies on sour fruit beverages by malolactic fermentation. IX Production of sour beverage from apple pomace. *Research Report of the Nagano State Lab. of Food Tech.*, 24:12-15.
- [208] Joshi, V.K. and Sandhu, D.K. 1996. Preparation and evaluation of an animal feed by product produced by solid-state fermentation of apple pomace. *Bioresour. Tech.*, 56(2,3):251-255.
- [209] Joshi, V.K., Kaushal, N.K., and Thakur, N.S. 1996. Apple pomace sauce—Development and quality of fresh and stored products. *J. of Food Sc. and Tech. (India)*, 35(5):414-417.
- [210] Ohsawa, K., Chinen, C., Takamami, S., Kuribayashi, T., and Kurokouchi, K. 1994. Studies on effective utilisation of carrot pomace I. Effective utilisation to bread. *Research Report of the Nagano State Laboratory of Food Tech.*, 22:24-28.
- [211] Ohsawa, K., Chinen, C., Takamami, S., Kuribayashi, T., and Kurokouchi, K. 1995. Studies on effective utilisation of carrot pomace II. Effective utilisation to cake, dressing and pickles. *Research Report of the Nagano State Lab. of Food Tech.*, 23:15-18.
- [212] Food Technology Abstracts, 1998. AFST (1), Mysore, India, 33:5.
- [213] Rasco, B.A. and McBurney, W.J. 1989. Human food product produced from dried distillers' spent cereal grains and solubles. *United States Patent US 4 828 846*.
- [214] Onodera, R., Kawamura, O., Inazawa, A., Izumi, T., Okuda, M., Katayama, H., and Yokoyama, M. 1996. Preparation of silages consisting of barley shochu distillery by-product and pulps (juice residues) of mandarin oranges and carrots. *Bulletin of the Faculty of Agri, Miyazaki University*, 43(2):145-150.
- [215] Roussos, S., Angeles-Aquihault, M.de los. Refugid Trejo-Hernandez, M. Del. Guime Perraud, I., Favela, E., Ramakrishna, M., Raimbault, M., and Viniestra-Gonzalez, G. 1995. Biotechnological management of coffee pulp, isolation, screening, characterization, selection of caffeine-degrading fungi and natural microflora present in coffee pulp and husk. *Appl. Microbiol. & Biotechnol.*, 42(5):756-762.
- [216] Mahadevaswamy, M. and Venkataraman, L.V. 1990. Integrated utilization of fruit processing wastes for biogas and fish production. *Biological Wastes*, 32(4):243-251.
- [217] Gandhi, V.M., Mukherjee, B., Iyer, V.J., and Cherian, K.M. 1997. Nutritional and toxicological evaluation of wild apricot pomace. *J. of Food Sc. and Tech. (India)*, 34(2):132-135.
- [218] Ghildyal, N.P. and Lonsane, B.K. 1990. Utilization of cassava fibrous residue for the manufacture of value added products: An economic alternative to waste treatment. *Process Biochem.*, 25(2):35-39.
- [219] Shah, G.H. and Masoodi, F.A. 1994. Studies on the utilization of wastes from apple processing plants. *Indian Food Packer (India)*, 48(5):47-52.
- [220] Carson, K.J., Collins, J.L., and Penfield, M.P. 1994. Unrefined, dried apple pomace as a potential food ingredient. *J. Food. Sci.*, 59(6):1213-1215.
- [221] Sudhakar, D.V. and Maini, S.B. 1995. Pectins from fruit processing waste—A review. *Indian Food Packer (India)*, 49(1):39-56.
- [222] Tressler, D.K. and Joslyn, M.A. 1961. *Fruit and vegetables juice processing technology*, pp. 1040. AVI, Westport, Connecticut.
- [223] Ihl, M., Astete, G., and Bifani, V. 1992. Preparation of pectins from apple pomace from the Araucania region of Chile with ethanol or AlCl₃. *Revista Espanola de Ciencia y Tecnologia de Alimentos*, 32(2):185-197.
- [224] Kunzek, H., Kruse, R., and Neumann, A. 1996. Low residue apple processing II. Manufacture of pectin and a low pectin material with cellular structure from stored wet apple pomace. *Flussiges Obst*, 63(6):314-319.
- [225] Contreras Lopez, A., Umeza Colla, H.M., Corona de la Torre, J., and Santamaria Diez, P. 1990. Extraction of pectin from apple waste produced in cider manufacture. *Research and Industry (India)*, 35(4):207-211.
- [226] Poonia, S., Yamadagini, R., and Dhawan, S.S. 1994. Studies of the utilization of citrus waste for pectin extraction. *Haryana J. of Horticultural Sc. (India)*, 23(1):28-32.
- [227] El-Nawawy, A.S. and Heikal, Y.A. 1996. Production of pectin pomace and recovery of leach lipids from orange peel. *J. of Food Engg.*, 28(3,4):341-347.
- [228] Tandon, D.K., Kaira, S.K., Singh, B.P., and Neelima Garg. 1991. Characteristics of pectin from mango fruit waste. *Indian Food Packer (India)*, 45(5):9-12.
- [229] Pedrera Islas, R., Aguilar Esperanza, E., and Vernon Carter, E.J. 1994. Obtaining pectins from solid wastes derived from mango (*Mangifera indica*) processing. *AIChE Symposium Series*, 90(300):36-41.
- [230] Martin-Cabrejas, M.A., Esteban, R.M., Lopez-Andreu, F.J., Waldron, K., and Selvendran, A.R. 1995. Dietary fibre content of pear and kiwi pomaces. *J. of Agril. and Food. Chem.*, 13(3):662-666.
- [231] Valiente, C., Arrigoni, E., Esteban, R.M., and Arzate, R. 1995. Grape pomace as a potential food fiber. *J. of Food Sc.*, 60(4):818-820.
- [232] Rodriguez de Sotillo, D., Hadley, M., and Holm, E.T. 1994. Potato peel waste: Stability and antioxidant activity of a freeze-dried extract. *J. Food Sc.*, 59(5):1031-1033.
- [233] Larrauri, J.A., Ruperez, P., and Calixto, F.S. 1996. Antioxidant activity of wine pomace. *Ame. J. of Eno. and Viti.*, 47(4):369-372.
- [234] Larrauri, J.A., Ruperez, P., and Calixto, F.S. 1997. Effect of drying temp. on the stability of polyphenols and antioxidant activity of red grape pomace peels. *J. Agri. and Food Chem.*, 45(4):1390-1393.
- [235] Peker, I. 1993. The obtaining of red colour pigment from red grape pomace. *Gida*, 18(4):269-272.
- [236] Kocoglu, A. and Algur, O.F. 1992. Test of media with vinasse for *Chlamydomonas reinhardtii* for possible reduction in vinasse pollution. *Bioresour. Technol.*, 42(1):1-5.
- [237] Mundigler, N., Herbinger, B., Berghofer, E., Schleining, G., and Grezes-kowiak, B. 1995. Detection of functional characteristics of a biogenous thermoplastic material. *Carbohydrate Polymers*, 26(4):271-278.
- [238] Mathias, A.L., Lopretti, M.J., and Rodrigues, A.E. 1995. Chemical and biological oxidation of *Pinus pinaster* lignin for the production of vanillin. *J. of Chem. Tech. and Biotech.*, 64(3):225-234.
- [239] Ramm, A., Baumann, G., and Gierschner, K. 1994. Waxes (including triterpenoids) in dried apple pomace and in dried residue after the extraction of pectin. *Industrielle Obst.—Und Gemuseverwertung*, 79(1):2-9.
- [240] Desai, A.J. and Pandey, S.N. 1971. Microbial degradation of cellulosic textiles. *J. of Scient. and Industr. Res. (India)*, 30:598-606.
- [241] Martinez, J.M., Granado, J.M., Montane, D., Salgado, J., and Fariol, X. 1995. Fractionation of residual lignocellulosics by dilute acid prehydrolysis and alkaline extraction: Application to almond shells. *Bioresour. Tech.*, 52(1):59-67.

- [242] Ghosh, P., Pamment, N.B., and Martin, W.R. 1982. Simultaneous saccharification and fermentation of cellulose: Effect of β -D-glucosidase activity and ethanol inhibition of cellulases. *Enz. Micro. Technol.*, 4:425-430.
- [243] Moo-Young, M. 1982. The waterloo SCP process: Direct conversion of cellulosic materials into proteinaceous foods. *Industry and Environment*, 5:30-31.
- [244] National Academy of Sciences. 1981. *Report on food, fuel, and fertilizer from organic wastes*. National Academy Press, Washington, D.C.
- [245] Crawford, D.L. and Crawford, R.L. 1980. Microbial degradation of lignin. *Enz. Micro. Technol.*, 2:11-22.
- [246] Kirk, T.K., Higuchi, T., and Chang, H.-M. 1980. Lignin bio-degradation: Summary and perspectives. In *Lignin biodegradation*, vol. 2, chap. 16, pp. 235-243. Kirk, T.K., Higuchi, T., and Chang, H.-M. Eds. CRC. Boca Raton, FL.
- [247] Amer, G.I. and Drew, S.W. 1980. Microbiology of lignin degradation. *Annu. Rep. Ferment Proc.*, 4:67-103.
- [248] Bungay, H.R. 1982. Biomass refining. *Science*, 218:643-646.
- [249] Busche, R.R., Ng, T.K., McDonald, C.C., and Hardy, R.W.F. 1983. Production of feedstock chemicals. *Science*, 219:733-739.
- [250] El-Nawawy, A.S., Fahmy, M., and Abdel Maick. 1966. Variables affecting maize cob pentosan conversion. In *Sixth chemical congress*. Cairo, Egypt.
- [251] El-Nawawy, A.S., Mahmoud, S.A.Z., Mashoor, W.M., and Ibrahim, E.M. 1974. Utilization of rice hulls for the production of microbial protein. In *Proceedings of 2nd Rice Conference*, Cairo, Egypt.
- [252] Wang, D.I.C., Cooney, C.L., Demuin, A.L., Gomez, R.F., and Sinskey, A.J. November, 1978. Degradation of cellulosic biomass and its subsequent utilization for the production of chemical feedstock. *Progress Report*, US Dept. of Energy Contract. EG-77-S-02-4198.
- [253] Wilke, C.R., Yang, R.D., and Von Stockar, V. 1976. Preliminary cost analysis for enzymatic hydrolysis of newsprint. *Biotechnol. Bioeng. Symp.*, 6:155-176.
- [254] Abbott, J.C. 1966. Unconventional Protein. In *Energy research conference*. Santa Barbara, California.
- [255] Wang, D.I.C. 26 August 1968. Protein from petroleum. *Chemical Engg.*, 75(18):100.
- [256] KISR. 1984. Kuwait Institute for Scientific Research. Report No. 1505. Kuwait.
- [257] Ruddle, C. 1982. Microbial oils and fats: An assessment of their commercial potential. *Prog. Indust. Microbiol.*, 16:119-206.
- [258] Solomons, G.L. 1976. Solvents from carbohydrates: Some economic considerations. *Proc. Biochem.*, 11(3):32-37.
- [259] Moore, C.A. 1977. Is agriculture a viable renewable raw materials resource? *Chemtech.*, 7:762-765.